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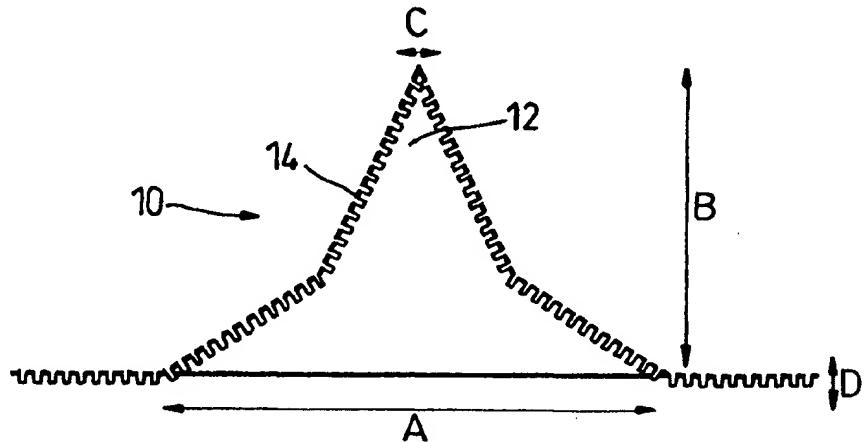
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(54) Title: TRANSFERRING MATERIALS INTO CELLS USING POROUS SILICON



(57) Abstract

The present invention relates to the use of porous silicon in the delivery of substances into cells. The porous silicon can be formed into micropiercers, microneedles and biostatic bullets for penetration of the cell. The control of the pore size and porosity of the porous silicon allows tuning of the bioactivity of the porous silicon. The porous silicon is also resorbable and is therefore resorbed from the cells without leaving any particles or being seen as a foreign body. The present invention also relates to the methods of manufacturing the porous silicon micropiercers, microneedles, microelectrodes, biostatic bullets, and precipitation of calcium phosphate on a bioactive substrate, and their advantages over known methods of delivering materials into cells.

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TRANSFERRING MATERIALS INTO CELLS USING POROUS SILICON

This invention relates to ways of transferring materials into cells,
5 and also to a microneedle array.

There are many times when it is necessary to transfer materials into cells, for example nucleic acids or nucleic acid constructs, such as vectors or plasmids, etc. have to be transferred into a cell for the purposes of 10 genetic manipulation. Furthermore, chemicals may also need to be transferred into cells, e.g. nucleotides or stains, and chemicals to affect the physiology of a cell. A number of chemical and mechanical processes have been developed to convey materials into cells. These techniques include:-

15

1. direct microinjection - a needle is inserted into a cell and material expelled through the needle;
2. electroporation - the cell membrane is made permeable to some 20 molecules by application of a high voltage shock;
3. biolistics - tungsten or gold particles are coated with the substance desired to be introduced and are shot into the cell;
- 25 4. calcium phosphate co-precipitation - cells absorb calcium phosphate, and if DNA/other material co-precipitates with the calcium phosphate it is also taken into the cell;
- 30 5. mediated transformation (via liposome, viral, or bacterial vectors); and

6. protoplast transformation.

An aim of one aspect of the present invention is to use a new
5 material to assist in the transfer of substances to cells.

An aim of another aspect of the invention is to provide an improved
way of providing small volumes of a substance.

10 Direct microinjection involves the insertion by a microneedle of
DNA directly into the nucleus of individual cells. A glass micropipette
linked to a micromanipulator is used to inject 10^{-8} - 10^{-7} μ l of material,
typically a solution of DNA fragments, into cell nuclei. "Hits" are almost
certain, given considerable operator expertise, but the technique is
15 laborious and cannot be applied to a large number of cells.

According to a first aspect the invention comprises a method of
transferring a substance into the cell.

20 Preferably resorbable or bioerodible porous silicon is used.

In one embodiment a microneedle that comprises at least a region
of porous silicon is used, or an array of such microneedles is used.

25 According to a second aspect the invention comprises a
microneedle (or microneedle array) comprising porous silicon.

According to a third aspect the invention comprises a vehicle for
transferring material into a cell, the vehicle comprising, at least in part,
30 porous silicon, and material to be transferred into the cell.

Preferably the porous silicon is resorbable. The vehicle may comprise a porous silicon biolistic bullet. The vehicle may comprise a substance which in use will co-precipitate with a co-precipitate substance 5 that is taken into the cell. The vehicle may comprise an electrically-conducting bioactive porous silicon electrode.

According to a fourth aspect the invention comprises the use of 10 porous silicon as a transfer medium for transferring materials into a living cell.

It has been discovered that porous silicon is biocompatible, and it has now been discovered that porous silicon can be corroded in, or resorbed into, a mammalian body without significant detrimental effect. 15 Porous silicon can be used to locate and substantially immobilise biological material (or any substance to be introduced into a cell), with the substance being free enough once in the cell to combine with cell DNA, or otherwise be released to have an effect.

20 It is known from PCT Patent Application No. WO 96/10630 to have an array of micromachined bulk silicon barbs or tips and to use them mechanically to pierce the plasma membrane of large numbers of cells simultaneously. This is more efficient than piercing a single cell with a single needle, which can result in a laborious operation if hundreds of 25 cells need to have material introduced into them. The tips of WO 96/10630 are, with hindsight, less effective at transferring material (e.g. DNA) into a pierced cell than they might be. It is, for example, proposed in that document to use surface tension forces between closely-spaced tips to hold biological material to be introduced into the cells in the spaces

between the tips, and to trap it between the tips (probes) and the substrate.

A proposal is discussed in US 5 262 128 which was published 5 in 1993 and purports to teach the man skilled in the art to make an array of silicon needles using the Liga Processes. It is believed that this document is non-enabling at its filing (and publication) date and is not prejudicial to the novelty of the present invention for that reason. In 1989 when the application was filed, and in 1993 when it was published, the 10 skilled man of ordinary expertise could not make very thin silicon needles having a central lumen as discussed in the document using the techniques discussed. The Liga Process is not suitable for manufacturing hollow needles in silicon, and does not enable sloping structures to be made.

15 US 5 457 041 discloses an array of solid needles made of silicon having ragged tips.

US 5 591 139 discloses a silicon microneedle that is formed in the plane of a silicon wafer.

20 An article on the Website www.gtri.gatech.edu/news/NEEDLES.html of Georgia Tech., apparently released on 22 June 1998, discusses making solid needle arrays from silicon, and expresses the hope to make hollow needles by an unknown, not yet identified, 25 technique.

US 4 969 468 discusses solid metal needles for electrical contact with nerves.

According to another aspect the invention comprises a cell-penetrating member or micropierce made of porous silicon.

The cell-penetrating member is adapted to have a substance to be
5 introduced into a cell carried by the porous silicon.

According to another aspect, the invention comprises a cell
penetrating member or a micropierce comprising at least a region of
porous silicon. Preferably the substance comprises DNA or RNA, a
10 fragment of DNA or RNA, or a construct of DNA or RNA.

The cell penetrating member or micropierce is preferably adapted
to have a substance to be introduced to a cell carried by porous silicon.

15 The porous silicon region is adapted to immobilise a substance
(e.g. DNA) in comparison with its mobility when provided with a bioinert
substance such as titanium. The porous silicon region is preferably at the
tip of the cell penetrating member or micropierce. The cell penetrating
member or micropierce may be a tip or barb, with no central lumen, or it
20 may be a needle with a central channel. The cell penetrating member or
micropierce may have a pore network extending from a reservoir or
channel to a substance delivery region provided on the surface of the cell
penetrating member or micropierce.

25 The cell penetrating member or micropierce may have a coating of
porous silicon, or it may be porous throughout its cross-section, at least at
its tip (or other substance delivery region if that is not the tip).
Substantially the whole exterior surface of the cell penetrating member or
micropierce that penetrates a cell in use may comprise porous silicon.

The cell penetrating member or micropiercer may be a bulk silicon microtip with a porous silicon coating.

An advantage of holding the substance to be introduced to the cell 5 at the tip of the cell penetrating member or micropiercer itself, instead of in channels/spaces between tips, is that the material is definitely introduced into the cell, and typically deeply into the cell. This may increase the success rate of the operation (in many cases introducing DNA into cells and stable uptake of the DNA/fragment is not statistically very 10 successful - a few percent may succeed, which is why so many cells have to be injected).

Instead of using porous silicon to immobilise the material on the tip/ensure at least some material is present on the tip, other holding means 15 may be used. For example, polycrystalline silicon can hold some substances at grain boundaries. The holding means may comprise a porous material.

It is known to immobilise DNA fragments in macroporous silicon 20 in the field of a flow-through genosensor (Advances in Genosensor Research. K.L. Beattie et al. Clin. Chem. 41, 700 (1995)).

An advantage of porous silicon is that its bioactivity can be tuned by controlling its pore size and porosity. It is therefore possible to create 25 a cell penetrating member or micropiercer with a porous tip with pores tailored to hold/immobilise a particular desired molecule or substance. Of course, the substance will not be so immobilised that at least some of the material cannot leave the tip when the tip is in the cell.

Porous silicon has another great advantage as the choice of material for a cell penetrating member or micropierce in that micromachining techniques for fabricating small scale devices from silicon exist, e.g. in the electronics industry.

5

It is known how to make a silicon structure porous (see for example US 5 348 618, the contents of which are hereby incorporated by reference).

10

An array of cell penetrating members or micropiercers may be provided. The array is preferably a two-dimensional array of $n \times m$ micropiercers. The micropiercers are preferably regularly disposed in a grid pattern, but they perhaps do not need to be.

15

It is also known to have an array of microtips for a completely different purpose - for field emission cathodes used in vacuum microelectronic applications. Here, a 5mm square silicon chip will typically contain about 500 microtips of pyramidal shape with tip widths of 50mm - 1 μm and heights of 10 - 100 μm , depending upon the manufacturing parameters chosen. With hindsight, these would be suitable for porosification and then use as micropiercers for transferring a substance into cells. It is also even known to have porous silicon pyramidal cathodes - e.g. Field emission from pyramidal cathodes covered in porous silicon. P.R. Wilshaw et al. J. Vac. Sci. Techn. B12,1 (1994);

20 Fabrication of Si field emitters by forming porous silicon. D. Kim et al. J. Vac. Sci. Tech. B14, 1906 (1996); and Porous silicon field emission cathode development. J.R. Jessing et al. J. Vac. Sci. Techn. B14, 1899 (1996). However, these are all in a totally different field, and none show a micropierce having held on it DNA, RNA, or any other substance to be

25 introduced into a cell.

According to a third aspect, the invention comprises a method of producing a micropierce device comprising manufacturing one or more micropierce projections, and providing substance holding means at or 5 near the tip of the projections.

Preferably the method comprises making at least a part of the projections porous. Preferably the method comprises making the tip of the projections porous or substantially the entire extent of the tips porous, 10 or providing a porous coating on the tip. Preferably the tip is made porous using an HF anodising technique.

According to another aspect, the invention comprises a method of transferring a material into a cell comprising associating the material with 15 a tip portion of a micropierce and piercing the cell with the micropierce.

Preferably the method comprises using porous silicon to locate the material at or near the tip portion.

20 According to a further aspect, the invention comprises a method of genetic manipulation of a cell comprising associating genetic material with a tip portion of a micropierce, piercing the cell with the micropierce to allow the genetic material to enter the cell. The genetic material may then be stably incorporated in the cell.

25

According to another aspect, the invention comprises a microneedle array comprising a plurality of needles extending away from a support, the needles each having fluid transport means adapted to transport fluid from their bases to their tips, and fluid supply means communicating with

the fluid transport means and adapted to supply fluid to be injected to the base of the needles.

Preferably the array of microneedles are made of silicon. It may 5 be micromachined, for example from a silicon wafer.

The fluid transport means may comprise a reservoir, which may extend under the needles. The support may have a lower portion, an upper portion, and a channel or reservoir extending between the upper and 10 lower portions, with the needles being provided in the upper portion and the fluid transport means extending to the reservoir or channel.

The fluid transport means may comprise a lumen, or macropore in each needle which may extend generally centrally of the needle through its 15 longitudinal extent. Alternatively, or additionally, the fluid transport means may comprise a pore or capillary network, such as a plurality of mesopores.

The array of needles may be provided on an integrated silicon chip, 20 which may also have a sensor provided on it, the sensor preferably enabling one to monitor in situ the transfection process. For example a photo emitter/detector may be used in association with light emitting markers (e.g. fluorescent) associated with the DNA. It may also be desirable to have a power supply and/or processing circuitry, and/or 25 control circuitry provided on the chip. Arrays of light emitting devices and photodetectors may enable the transfection process to be monitored under high spatial resolution.

According to another aspect, the invention comprises a method of 30 manufacturing a microneedle, or a microneedle array, the method

comprising taking a bulk silicon wafer and creating a needle or an array of needles; and creating fluid transfer means extending from the base of the or each needle to its tip.

5 Preferably, the method comprises providing a network of pores from the base of the or each needle to its tip. The pores may be macropores or mesopores, or for some applications they may even be micropores (but macropores are preferred).

10 The or each needle may be created using photolithographic techniques such as anisotropic etching and photo-resist lithographic techniques.

15 The silicon substrate may be an n-type substrate with a resistivity in the range of 0.1-10Ωcm.

The needle or needle array may be planarised, for example by use of a non-conducting mask.

20 The planarised array may then be treated so as to expose just the tips, for example by using an oxygen plasma treatment and an HF dip to expose the tips alone. The planarised array may also be in-filled. The tips can then be anodised to create the pores from the tip to the wafer back surface. The wafer, provided with an array of tips, may then be 25 bonded to another backing member, which may be shaped so as to define a channel or reservoir between the tip-carrying wafer and the backing member.

According to a further aspect the invention comprises a vehicle for transferring material into a cell, the vehicle comprising at least in part resorbable material.

5 Preferably the vehicle comprises resorbable silicon, such as porous silicon, or polycrystalline silicon. The whole of the vehicle may be made of the resorbable material, or only part of it. The vehicle may comprise bioactive silicon. (By "resorbable" it is meant that the material is corroded/absorbed/eroded/ or otherwise disappears when in situ in
10 physiological fluids. By "bioactive" it is meant that the material can induce the deposition of calcium phosphate precipitates on its surface under physiological conditions (when in body fluids)).

If the vehicle is retained in the cell it will be
15 adsorbed/corroded/eroded or resorbed, or partially resorbed, and be less of an irritation/foreign body to the cell in due course.

The resorbable silicon/other material may be used in a biolistics technique. For example the vehicle for transferring material into a cell
20 may be a ballistic bullet.

The vehicle for transferring material into the cell may comprise a ballistic bullet comprising porous silicon.

25 The bullet may have a substance to be introduced into a cell adhered to it. The bullet may be impregnated with material (e.g. DNA material). It may be substantially saturated with material. The bullet may comprise a submicron silicon particle. The silicon particle may be rendered porous by stain etching techniques. The particle is preferably
30 mesoporous.

A resorbable ballistic bullet would not leave behind in the cell a particle, as do gold or tungsten ballistic bullets. The bullet need not be porous all of the way through - it may have a porous coating. The 5 resorbable bullet need not necessarily be made of porous silicon, or of silicon at all, but porous silicon has been identified as an especially suitable material.

According to another aspect, the invention provides a method of 10 transferring material into a cell comprising the steps of shooting a vehicle carrying said material into the cell.

Preferably the vehicle is the vehicle as hereinabove defined. Preferably the bullet is shot into the cell by means of a pressurised gas, 15 for example helium.

The process of biological biolistics is often used where more standard techniques do not work. Resorbable impregnated materials, such as porous silicon offer biocompatible advantages over corrosion-resistant 20 bulk metal materials.

According to a further aspect the present invention provides a method of making a vehicle for transferring material into a cell comprising the steps of rendering the vehicle at least partially porous and 25 introducing to the vehicle the material to be transferred to the cell.

Preferably the vehicle comprises a silicon bullet, most preferably a submicron silicon particle, which may be rendered porous, preferably mesoporous by stain etching techniques. The bullet may have the material

to be introduced to the cell, adhered to it or alternatively it may be impregnated with the material.

5 The vehicle may be a submicron particle and the material to be transferred to the cell may be co-precipitated (with a precipitate substance) using the vehicle as a nucleation site.

10 The vehicle for transferring material into a cell may comprise bioactive silicon. The vehicle may have associated with it material to be transferred in a form adapted to co-precipitate with a substance which is taken up by cells. The co-precipitate may be a calcium phosphate precipitate.

15 According to another aspect the invention comprises a method of introducing material into a cell comprising associating the material with a silicon particle, precipitating calcium phosphate onto the particle to form a calcium phosphate/silicon particle combined particle, and arranging for the cell to uptake the calcium phosphate/silicon particle combination.

20 In the technique of electroporation the cell membrane can be made permeable by exposing cells to a brief electric shock of very high voltage. Low porosity bioactive silicon is electrically conducting and is suitably developed as an intimate coupling matrix for adherent mammalian cells growing on microelectrode arrays.

25

By having bioactive silicon, e.g. porous silicon or polycrystalline silicon, as one or both electrodes in electroporation apparatus it is envisaged that better DNA transfer takes place.

According to a further aspect the invention comprises a method of electroporation comprising providing an electrically conducting bioactive silicon electrode.

5 Preferably the method comprises growing cells on the electrode. The method may comprise providing an array of bioactive silicon electrodes, possibly with cells grown on them. The electrode, or electrodes, may be coated with porous silicon or may be of porous silicon throughout their cross-section, at least at a region of their height.

10 According to a further aspect the invention comprises electroporosis apparatus comprising a bioactive electrode. Preferably the electrode is bioactive silicon, most preferably porous silicon. An array of electrodes, or microelectrodes, may be provided.

15 The invention may also reside in the use of bioactive silicon, preferably porous silicon, in the preparation of apparatus for the introduction of materials into cells.

20 It may be helpful to clarify that bioactive materials are a class of materials which when *in vivo* elicit a specific biological response that results in the formation of a bond between living tissue and that material. Bioactive materials are also referred to as surface reactive biomaterials. Resorbable materials are materials which are designed to disappear or
25 degrade gradually over time *in vivo*, and they may or may not be replaced with tissue. Bioerodable materials are materials which erode *in vivo*, with the material possibly being absorbed by cells, or possibly not being absorbed. A bioinert material is a material that elicits no local gross biological response *in vivo*.

Several embodiments of the present invention will now be described by means of example only with reference to the Figures, in which:-

5 **Figure 1** shows a partially porosified silicon microtip;

Figure 2 shows a silicon microneedle array;

10 **Figure 3** shows the silicon microneedle array of **Figure 2** having a macroporous network running from the tip to an underlying reservoir;

Figure 4 shows a porous silicon bullet impregnated with DNA;

15 **Figure 5** shows a porous silicon core impregnated with DNA and surrounded by calcium phosphate;

Figure 6 illustrates the electroporation technique of the present invention; and

20 **Figures 7 and 8** show SIMS plots demonstrating the affinity of DNA for porous silicon and that it can be released from a porous silicon surface.

25 **Figure 1** shows a micropierce 10 in the form of a microtip 12 having a base width A of 50 μ m and a height B of 100 μ m and a tip width C of 0.5 μ m. The surface of the microtip 12 is coated with porous silicon 14 having a depth D of 0.1 μ m.

In use, the porous silicon coating 14 immobilises the substance to be delivered to the cell (e.g. DNA/RNA) on the tip itself, which increases the chances of the immobilised substance on the tip being introduced into the living cell.

5

The pore size and porosity of the porous silicon coating can be controlled to tune the bioactivity of the microtip 12. By controlling the pore size and porosity of the porous silicon, we can make particular molecules come off it more, or less, readily. We may leave the microtips 10 inside a cell for a predetermined time to allow molecules to disassociate themselves from the porous silicon.

Figure 2 shows an array 20 of silicon microneedles 22 extending away from a silicon support, or back, member 24. The microneedles 22 15 have porous silicon microtips 26 and a central lumen 28 communicating between the microtips 26 and a reservoir 30 defined between an upper member 32, provided with the microneedles 22 and the back, support, member 24. The back member 24 is of bulk silicon.

20

Figure 3 shows an array 33 of silicon microneedles 34 that is similar to that of Figure 2. The principal difference between the arrays shown in Figures 2 and 3 is that the microneedles 34 shown in Figure 3 are not provided with a central lumen 28. Instead the array 33 of silicon microneedles 34 in Figure 3 is provided with a mesoporous 25 network 36 which extends from the microtips of the microneedles to the reservoir 30', allowing fluid communication between the reservoir 30' and the microtips.

In use, the substance to be delivered to cells is provided to the 30 porous silicon microtips 22,34 from the reservoir 30,30' through the

central lumens 28 or the mesoporous network 36. The substance is then held by the porous silicon microtips ready for introduction into a cell.

The material to be introduced into the cells may be pumped into the 5 reservoir 30, 30', and out through the lumens 28 or porous network by a pump, not shown (but arrow 39 indicates the pump delivering liquid to the reservoir).

All or part of the silicon surfaces within the final structure may be 10 treated in such a way as to modify their interaction with biological systems. This might be achieved by forming a layer of porous silicon on the surface. Such a layer could be formed by either an electrochemical anodisation process or possibly by immersing the structure into a stain etching solution such as a mixture of hydrofluoric acid and nitric acid.

15

Figure 4 shows a ballistic bullet 40 comprising a submicron silicon particle rendered mesoporous by stain etching.

In use, the bullet 40 is impregnated with the substance to be 20 introduced into a cell and is shot into the cell using pressurised helium. As the porous silicon is a resorbable material, it will be preferably fully resorbed, and at least partially resorbed, by the cell that it entered, and thus comprises less of a foreign body than known ballistic bullets such as gold or tungsten which leave particles of metal in the cell.

25

Figure 5 shows a porous silicon core 50 impregnated with a substance to be introduced into a cell (e.g. DNA/RNA) and calcium phosphate precipitate 52 formed around the core 50. The calcium phosphate 52 is co-precipitated with DNA/RNA, so that a genetic 30 material/calcium phosphate layer surrounds the bioactive silicon core 50.

The bioactive silicon core locally induces calcium phosphate supersaturisation. It may be possible to place a bioactive silicon core next to a cell/against the wall of a cell, and co-precipitate DNA/Ca(PO₄)₂ against the core and against the wall of the cell. If the core is 5 phagocytosed it can be resorbed.

The core 50 need not have DNA/RNA/any active substance on it - it may simply serve as a good nucleation site for co-precipitation of DNA/Ca(PO₄)₂.

10

It is known to use glass beads as a nucleation site for calcium phosphate co-precipitation DNA transfection - see for example the paper by Watson and Latchman in "Methods (San Diego) 1996 10(3), 289-291 (Eng).

15

It will be appreciated that micropores are pores with a diameter of 2 nm or less; mesopores have a diameter of 2nm - 50 nm; and macropores have a diameter of 50 nm or more.

20

It has also been realised that it is possible to improve the efficiency of the introduction of materials to cells in an electroporation technique, as shown in Figure 6, using porous silicon, preferably mesoporous silicon (but macroporous and microporous silicon are also useful).

25

The use of a porous silicon (or porous other bioactive material, or bioactive polycrystalline silicon) electrode 60,61 achieves better performance in electroporation. Because the electrode is bioactive, instead of being bioinert, cells (typically animal cells) have an affinity to it and are localised on its surface.

30

Low porosity (50% or less, or 30% or less, or 10% or less) bioactive silicon is electrically conducting and is a suitable intimate complex matrix for adherent mammalian cells 62, which may grow on a microelectrode array 60,61. Thus, it is possible to grow mammalian cells 5 on bioactive porous silicon electrodes and then introduce DNA (or other substances) into the cells by using electroporation, with the substrate upon which the cells are grown being an electrode, or even both electrodes 60,61, of the electroporation apparatus. This has advantages in handling the cells, and achieves a better efficiency rate of DNA 10 introduction than solely having the cells suspended in a liquid medium 63.

The fact that porous silicon is resorbable/erodable in vivo in mammals has been proved by the inventors, and this underpins some aspects of the invention. The fact that silicon can be made bioactive 15 underpins other aspects of the invention.

Figure 7 shows a SIMS plot (Secondary Ion Mass Spectroscopy) showing the concentration of nitrogen with depth in a sheet of porous silicon. DNA is rich in nitrogen, and detecting high nitrogen levels in the 20 porous silicon is a measure of how much DNA is present. Plot 70 shows the "aged" porous silicon sheet analysed for nitrogen, with no DNA added to the surface of the sheet. The background level of nitrogen depends on the type of porous silicon film and its "age" - the duration of storage in ambient air. Plot 72 shows the analysis of the porous silicon sheet after a 25 single drop of water has been applied to the surface of the porous silicon sheet. There was 1ng per μ litre of DNA in the drop of water. The DNA solution drop was dried at 50°C before the sheet was analysed. Plot 74 shows the amount of nitrogen in the porous silicon when the same 1ng per μ litre of DNA in water drop is applied to the sheet and dried, and then 30 the sheet is washed in deionised water at 50°C.

As will be seen, there is far more nitrogen shown in plot 72 than in plot 70, showing that the DNA is being detected by the test. Plot 74 shows that the washing step removed some, but not all, of the DNA - that 5 some of the DNA was probably partially immobilised on the porous silicon, to be released later (during washing).

Figure 8 shows equivalent SIMS plots for the same layer after pure water treatment. "Aged" porous silicon has been stored in ambient air 10 and has acquired a background level of nitrogen due to adsorption of nitrous oxides and ammonia - common trace pollutant gases. Plot 80 shows aged porous silicon with no DNA, plot 82 aged porous silicon with a water droplet deposit (no DNA solution), and plot 84 shows again for comparison the analysis of aged porous silicon with 1ng/ μ litre of DNA in 15 solution added and dried at 50°C.

The SIMS data shown in Figures 7 and 8 demonstrate that porous silicon can reversibly bind DNA.

20 The invention can perhaps be thought of as using porous silicon (or perhaps polycrystalline silicon) as an inorganic vector for transporting/transferring material into a living cell.

CLAIMS

1. A method of transferring a substance into a cell comprising using porous silicon for conveying the substance into the cell.
- 5 2. A method according to claim 1 which comprises using a resorbable or bioerodable porous silicon.
- 10 3. A method according to claim 1 or claim 2 comprising using a microneedle that comprises at least a region of porous silicon.
4. A method according to claim 3 comprising having at least the tip of the needle provided with porous silicon.
- 15 5. A method according to any preceding claim which comprises using a microneedle (i) having a coating of porous silicon or (ii) made substantially completely of porous silicon; at least for a portion of the length of the microneedle.
- 20 6. A method according to any preceding claim comprising using an array of microneedles which comprise at least in part porous silicon.
7. A method according to any preceding claim which comprises using a microneedle or microneedle array, in which the microneedles are hollow and comprise porous silicon, the substance being provided in the hollow or transferred through the hollow.
- 25 8. A method according to any preceding claim in which a microneedle array is used, the array comprising a plurality of needles extending away from a support, the needles comprising substantially completely, or at
- 30

least in part, porous silicon, and each needle having fluid transfer means adapted to transport fluid from their bases to their tips, and fluid supply means communicating with the fluid transport means and supplying fluid to be injected to the base of the needles.

5

9. A method according to any of claims 3 to 8 in which the needle has no central lumen and comprises a micropiercer, porous silicon being provided on the needle, the porous silicon holding the substance to be conveyed.

10

10. A method according to any one of claims 1 to 9 comprising providing a needle or needles with a pore network extending from a reservoir or channel to a substance delivery region provided at the surface of the needle.

15

11. A method according to any preceding claim in which the porous silicon immobilises the substance in comparison with its mobility when provided on bioinert materials such as bulk titanium.

20

12. A method according to claim 1 or claim 2 which comprises using a porous silicon biolistic bullet.

25

13. A method according to claim 1 or claim 2 which comprises using porous silicon having the substance associated with it, and providing the porous silicon in a form adapted to co-precipitate with another substance to form a co-precipitate which is taken into the cell.

14. A method according to claim 13 comprising using calcium phosphate as the co-precipitate.

30

15. A method according to claim 1 or claim 2 comprising using an electrically bioactive electrode that comprises at least in part porous silicon, and using electroporation to convey the substance into the cells.
- 5 16. A method according to claim 15 in which the cells adhere to the electrode.
- 10 17. A method according to any preceding claim in which the substance comprises DNA or RNA, a fragment of DNA or RNA, or a construct of DNA or RNA.
18. A microneedle comprising porous silicon.
19. A microneedle according to claim 18 which has a duct.
- 15 20. A microneedle according to claim 19 in which the duct extends from the base region of the needle to the tip of the needle.
- 20 21. A microneedle according to any one of claims 18 to 20 in which at least a portion of the needle is made substantially completely of porous silicon.
22. A microneedle according to any of claims 18 to 21 in which at least a part of the needle comprises a coating or surface layer of porous silicon.
- 25 23. A microneedle according to any one of claims 18 to 22 in which a porous or capillary network is provided.
- 30 24. A needle array according to any one of claims 18 to 23 which further comprises a substance adapted to be conveyed into a cell.

25. A needle according to claim 24 in which the substance is carried by, or held on the needle, by the porous silicon.

5 26. A needle according to any one of claims 18 to 25 which is resorbable or bioabsorbable, or at least part of which is resorbable or bioabsorbable.

10 27. An array of microneedles extending away from a support, in which the microneedles are in accordance with any of claims 18 to 26.

28. A vehicle for transferring material into a cell, the vehicle comprising, at least in part, porous silicon, and material to be transferred into the cell.

15 29. A vehicle according to claim 28 in which the porous silicon is resorbable.

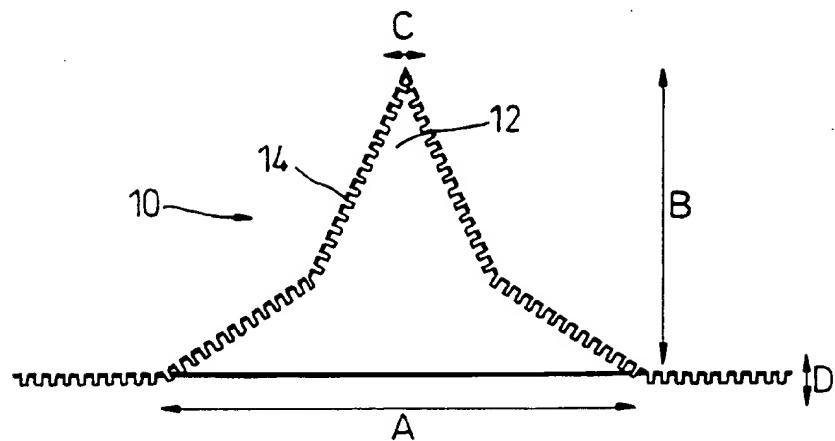
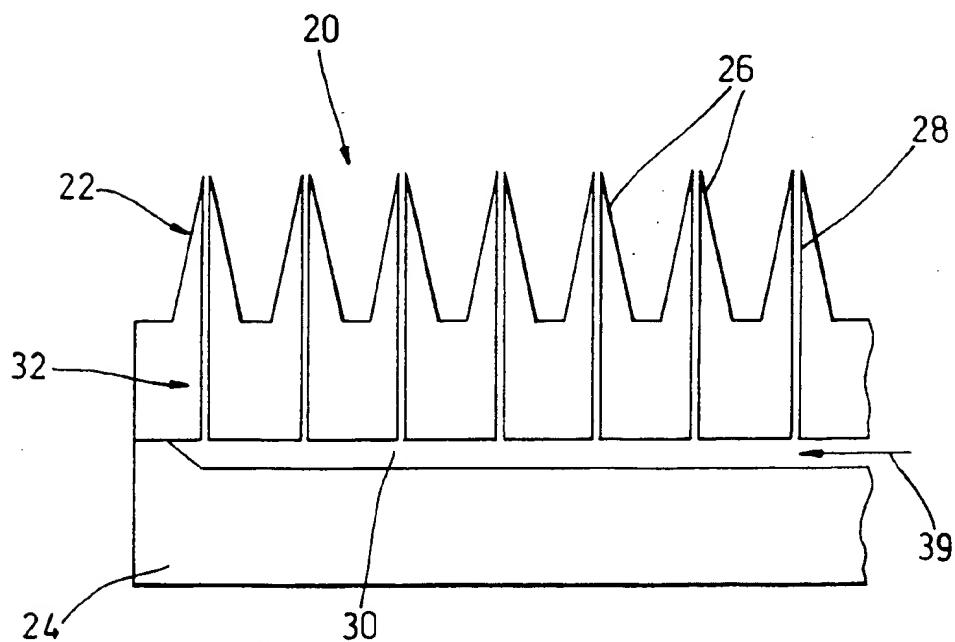
30. A vehicle according to claim 28 or claim 29 which comprises a porous silicon biostatic bullet.

20 31. A vehicle according to claim 28 or claim 29 in which the vehicle comprises a substance which in use will co-precipitate with a co-precipitate substance that is taken into the cell.

25 32. A vehicle according to claim 28 or claim 29 which comprises an electrically-conducting bioactive porous silicon electrode.

30 33. The use of porous silicon as a transfer medium for transferring materials into a living cell.

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*Fig. 1**Fig. 2*

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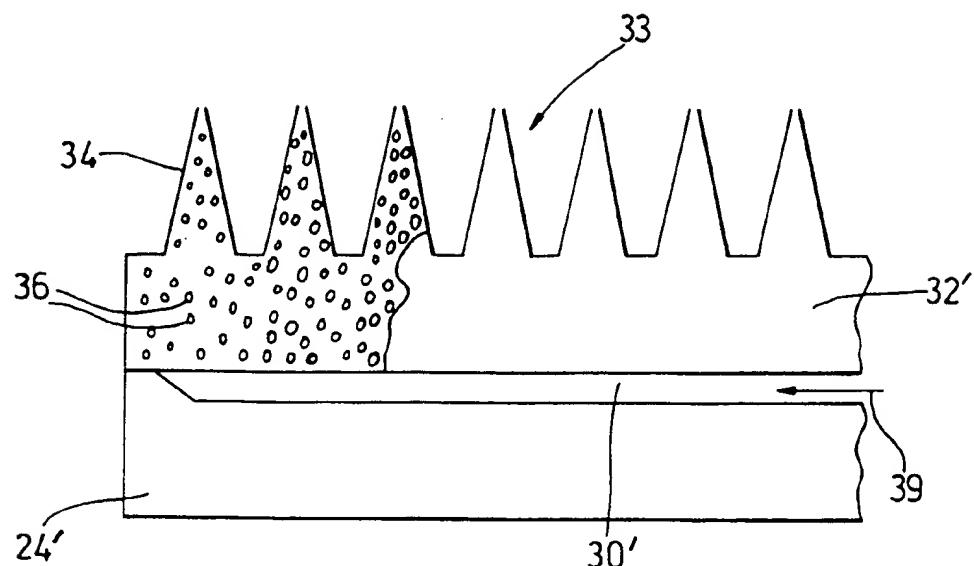


Fig. 3

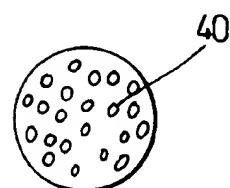


Fig. 4

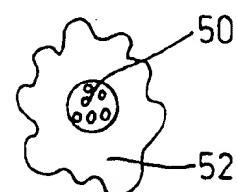
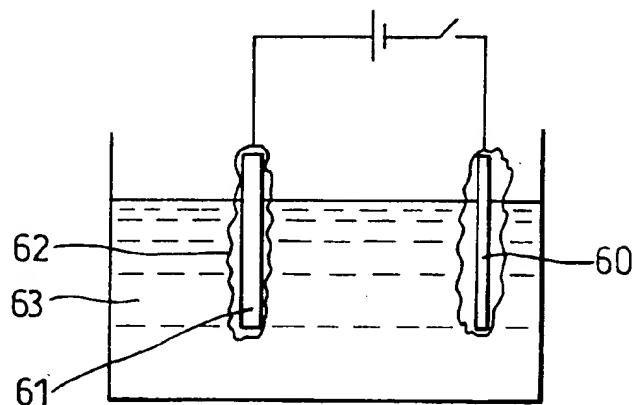
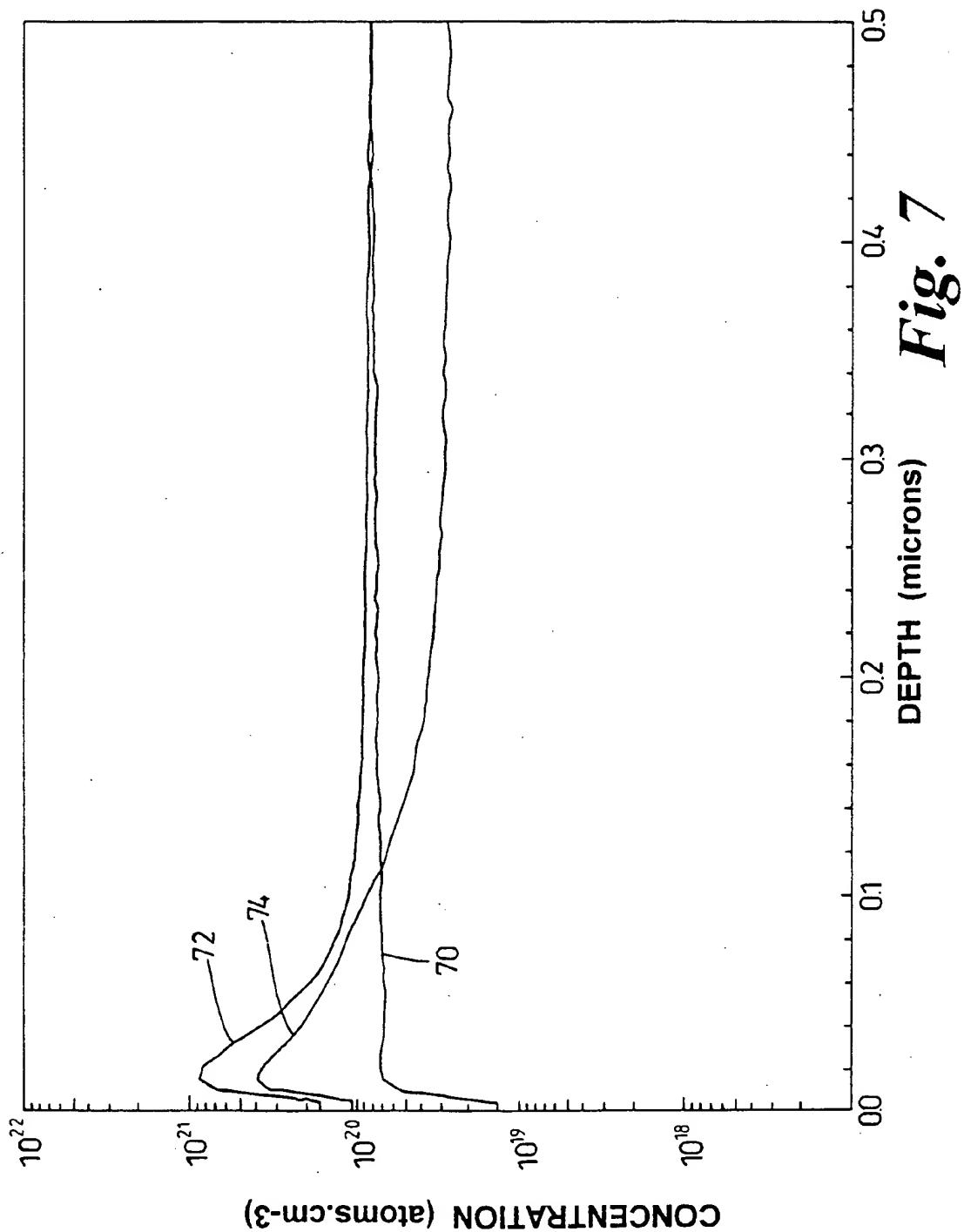


Fig. 5

Fig. 6

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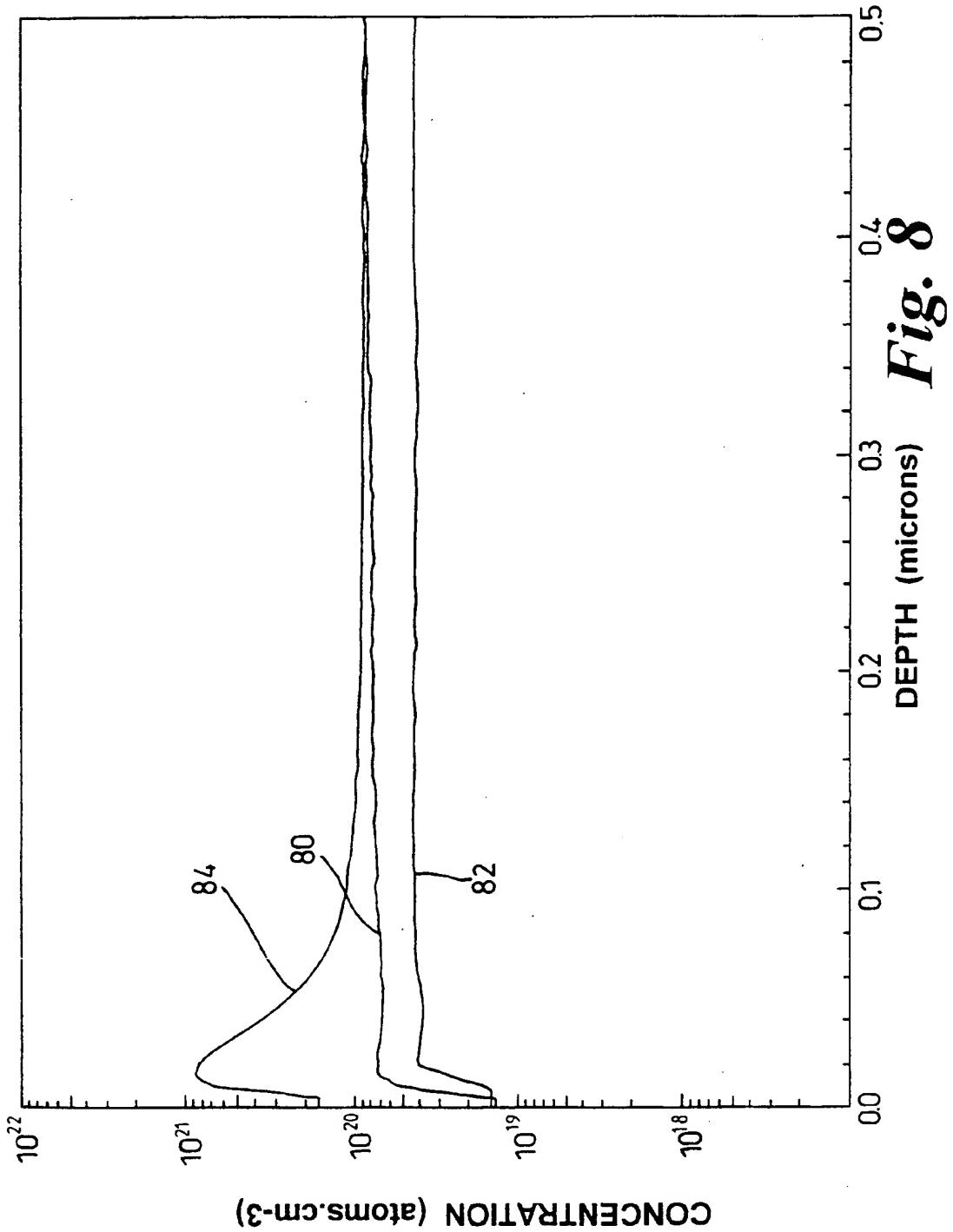


Fig. 8

INTERNATIONAL SEARCH REPORT

Interr. Application No

PCT/GB 99/02383

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 7 C12M3/00 C12N15/89 C12N15/87

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12M C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 97 06101 A (SECR DEFENCE ;CANHAM TREVOR LEIGH (GB)) 20 February 1997 (1997-02-20) abstract ---	1-33
A	US 5 457 041 A (GINAVEN ROBERT O ET AL) 10 October 1995 (1995-10-10) cited in the application abstract ---	1-33
A	BEATTIE K L ET AL: "ADVANCES IN GENOSENSOR RESEARCH" CLINICAL CHEMISTRY, vol. 41, no. 5, 1 May 1995 (1995-05-01), pages 700-706, XP000605273 ISSN: 0009-9147 cited in the application page 704 -page 706 ---	1-33
		-/-

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
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Date of the actual completion of the international search

3 November 1999

Date of mailing of the international search report

10/11/1999

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INTERNATIONAL SEARCH REPORT

Interr. Application No

PCT/GB 99/02383

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 96 10630 A (UNIV RUTGERS) 11 April 1996 (1996-04-11) cited in the application abstract ----	1-33
X	PATENT ABSTRACTS OF JAPAN vol. 018, no. 250 (P-1736), 12 May 1994 (1994-05-12) & JP 06 034361 A (CANON INC), 8 February 1994 (1994-02-08) abstract ----	18, 21, 22
A	SANFORD J C ET AL: "OPTIMIZING THE BIOLISTIC PROCESS FOR DIFFERENT BIOLOGICAL APPLICATIONS" METHODS IN ENZYMOLOGY, vol. 217, 1993, pages 483-509, XP002912840 ISSN: 0076-6879 table I ----	12
A	WO 91 05519 A (US ARMY) 2 May 1991 (1991-05-02) cited in the application abstract; figure 4 ----	7
A	WO 92 01802 A (GILMORE JAMES ;BRINEGAR CHRIS (US); JOHNSON MICHAEL (US); WALKER N) 6 February 1992 (1992-02-06) abstract ----	29

INTERNATIONAL SEARCH REPORT

Information on patent family members

Internat'l Application No

PCT/GB 99/02383

Patent document cited in search report	Publication date	Patent family member(s)			Publication date
WO 9706101	A 20-02-1997	GB 2303847 A			05-03-1997
		AU 6626496 A			05-03-1997
		CA 2228426 A			20-02-1997
		EP 0842113 A			20-05-1998
		GB 2317885 A			08-04-1998
		JP 11510469 T			14-09-1999
US 5457041	A 10-10-1995	NONE			
WO 9610630	A 11-04-1996	AU 3758595 A			26-04-1996
		CA 2201315 A			11-04-1996
		EP 0782614 A			09-07-1997
JP 06034361	A 08-02-1994	NONE			
WO 9105519	A 02-05-1991	US 5262128 A			16-11-1993
		AU 6640190 A			16-05-1991
		EP 0497885 A			12-08-1992
WO 9201802	A 06-02-1992	AU 8418091 A			18-02-1992
		US 5219746 A			15-06-1993